REVIEWED
By Celena Shrum at 9:37 am, May 06, 2021

5/6/2021

Worklist: 4953

LAB_CASE	ITEM	ITEM_TYPE	DESCRIPTION
M2021-1056	2	UCK	AM 6 Urine GHB

SC

AM 6: Urine GHB Screening Extraction

Extraction Date: 5/5/21 Analyst: Sarah Collins

Mobile phase A: 0.1% Formic Acid in Water **Mobile phase B:** 0.1% Formic Acid in MeOH
Blank Urine Lot: POC031319 **Column**: Phenomenex Phenyl Hexyl (4.6x50mm, 2.6um)
LCMS-QQQ ID: 069901
GHB-D6: FE07031801Cerilliant
GHB: FE04111903 Cerilliant

Pre-Analytic:

☒ 1. *Positive Control Working Solution Preparation Instructions:*
- *Working Solution:* Preparation of 200,000 ng/mL Positive Control Working Solution: Add 200µL of GHB 1 mg/mL stock solution to 800µL negative urine.
- Preparation of 10,000 ng/mL Positive Control: Add 10µL of GHB 20,000 ng/mL working solution to 190 µL negative urine.

☒ 2. Check levels of mobile phases and needle wash refill as needed. Ensure waste is not full.

☒ 3. Ensure correct column is installed and begin mobile phase flow allow to equilibrate ~ 30 minutes.

Analytic:

☒ 1. Remove working solutions, controls, and samples from cold storage.

☒ 2. Label centrifuge tubes for positive control, negative control and case samples.

☒ 3. Label ALS or LCMS vials for positive control, negative control, and case samples. Place insert in all vials.

☒ 4. Place on tube rocker at ambient temp for approx. 10 minutes.

☒ 5. Pipette positive and negative controls (for negative control, 200 µL urine will be added to the appropriate tube). Add 200µL urine to each centrifuge tube for case samples.

☒ 6. Add 100µL of the GHB-D6 Internal Standard Working Solution to each tube.

☒ 7. Add 900µL of 0.1% formic acid in methanol to each tube. Vortex.

☒ 8. Centrifuge at ~3400 rpm for 15 minutes.

☒ 9. Add 100µL 0.1% formic acid in water to each vial insert.

☒ 10. Transfer 10µL of sample from each centrifuge tube to the corresponding vial insert (avoid disturbing the pellet at the bottom). Vortex.

Post-Analytic

☒ 1. Open quantitation software and create a new quantitation batch.

☒ 2. Using the positive control, a 1-point calibration curve will be established. The curve will be set to linear, non-weighted and origin set to force.

☒ 3. If a sample gives a response that is greater than 10,000 ng/mL, a statement on the report will be included saying that preliminary testing indicated a possible presence of an elevated level of GHB and that it is recommended that the sample be sent to a private lab for quantitation. If a sample gives a response between 7,000 and 10,000 ng/mL, an inconclusive statement can be added to the report.

☒ 4. The S/N for samples and controls at and over 10,000 ng/mL must be 5 or greater

☒ 5. Case samples and negative controls will generally be considered negative if the calculated concentration is less than 7,000 ng/mL.

☒ 6. Central File Packet to include: LIMS Worklist, Method Checklist, Working solution prep sheet(s), Calibration and Control Reports

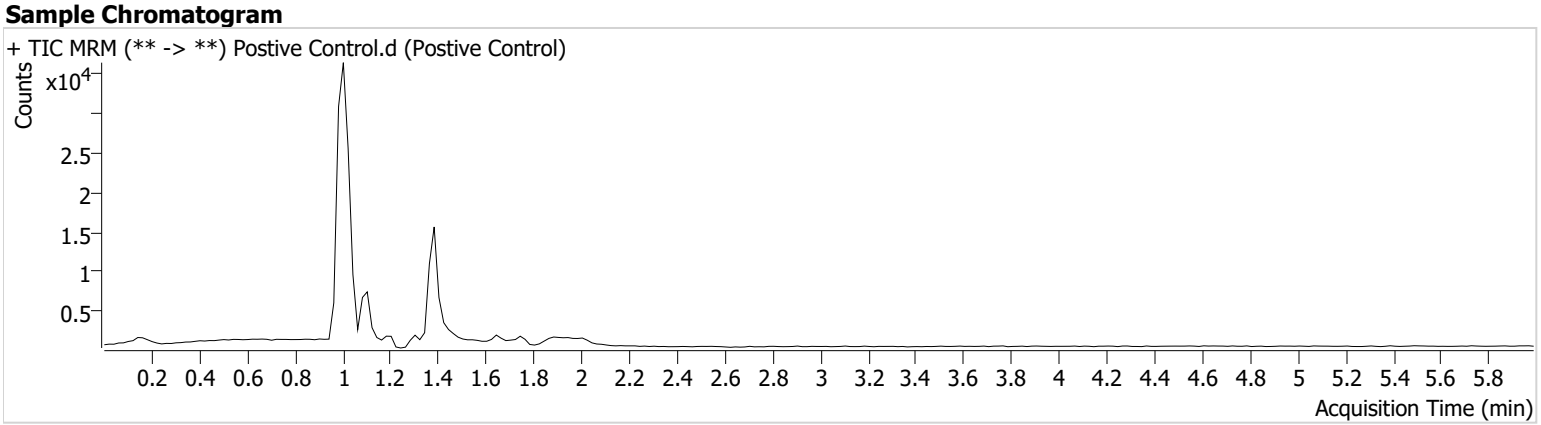
COMMENTS: The working solutions for the GHB positive control and internal standard were made using the lots above on 5/4/21.

AM #6 GHB Screen Results

Batch results D:\MassHunter\Data\2021\GHB\050521 GHB SC\QuantResults\GHB.batch.bin
Calibration Last Update 5/6/2021 9:00:56 AM

Instrument	Falco	**Data File**	Postive Control.d
Type	Cal	**Sample**	Postive Control
Acq. Method	GHB urine screen.m	**Operator**	Sarah Collins
Sample Position	P1-A2	**Comment**	
Injection Volume	2.5		
Acq. Date-Time	5/5/2021 2:34:31 PM		
Sample Info.			

Sample Chromatogram



Name	RT	Resp.	S/N	S/N	ISTD Resp.	Calc. Conc.
GHB	1.386	18226	83.79	84.90	10014	10000.0000

AM #6 GHB Screen Results

Batch results	D:\MassHunter\Data\2021\GHB\050521 GHB SC\QuantResults\GHB.batch.bin
Calibration Last Update	5/6/2021 9:00:56 AM

Instrument	Falco	**Data File**	Negative Control.d
Type	Sample	**Sample**	Negative Control
Acq. Method	GHB urine screen.m	**Operator**	Sarah Collins
Sample Position	P1-A1	**Comment**	
Injection Volume	2.5		
Acq. Date-Time	5/5/2021 2:27:56 PM		
Sample Info.			

Sample Chromatogram